Effect of Premium Motor Spirit (Petrol) on Lethality of *Tympanotonus Fuscatus* after Acute Exposure

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ABSTRACT

Tympanotonus fuscatus acclimated to laboratory conditions for seven days were exposed to different concentrations (30, 60, 90, 120 and 150ml/L) of petrol and a control for 96 hours to examine the acute effect of the petrol on Tympanotonus fuscatus. The response of the Tympanotonus fuscatus in the toxicant showed that mortality was concentration and time dependent. The 48^{th} and 96^{th} hour LC_{50} were 127.36 and 34.12ml/L respectively. While the 48^{th} and 96^{th} hour LC_{99} were 396.82 and 126.64ml/L respectively. The probit equations were however significant at 48, 72 and 96^{th} hour. The probit equations were: Y = -1.10 + 0.1X, Y = -0.91 + 0.014X and Y = -0.86 + 0.25X respectively. The mean lethal time (MLT₅₀) and their associated confidence limits were: 30mg/L, 77.13 (38.50-92.11), 60 mg/L, 61.64 (30.13-86.41), 90mg/L, 68.09 (29.35-75.58), 120mg/L, 44.71 (22.98-58.30) and 150 mg/L, 43.17 (16.95-56.30). The results obtained indicated that petrol is very toxic to Tympanotonus fuscatus. Therefore adequate measure should be put in place to check avoidable spill in the environment, more especially the aquatic environment which is the natural habitat of many susceptible aquatic species.

Keywords: Premium Motor Spirit, Petrol, Lethality, Tympanotonus Fuscatus

INTRODUCTION

The aquatic ecosystem is continuously subjected to changes in quality due to the introduction of substances of diverse characteristics arising from man's activities (Oluah, 2001). Oil exploration, exploitation, transportation and storage of crude oil have greatly affected the marine environment (Doertter, 1992). Petroleum hydrocarbons are the major

contaminants of the estuarine and coastal environments (Windows *et al.*,1982), and are quantitatively the most important constituents of petroleum. They arise from natural as well as anthropogenic sources (Law and Biscaya, 1994; Medeiros *et al.*, 2005). The control of such pollution problems in the aquatic environment is difficult because of the large number of input sources and their geographic dispersions (Howard et al, 2009). Studies have also shown that petroleum hydrocarbons mixes with water and penetrates to the underlying sediments (Carbioch *et al.*, 1977; Patin, 1999) and therefore may constitute a problem to benthic organisms which are particularly vulnerable to oil spills and forage the bottom sediments into most pollutants (Sprague *et al.*, 1981).

Petroleum (crude oil) poses a great risk to aquatic organisms which are sources of protein for the coastal dwellers in the Niger Delta (Doertter, 1992). Such risk includes: mortality (Ewa-Oboh and Otogo, 2009), stress (ATSDR, 1999), changes in species composition, low abundance, loss of species and tainting (Widdows *et al.*, 1982), changes in physiological and biochemical response (Lopes et al., 2001; Jee and Kang, 2005).

Crude oil and its products vary considerably in their toxicity and sensitivity of the organism to the product (Lopes et al., 2001). Due to the problem of crude oil toxicity examination resulting from batch differences, investigations are directed towards testing the biological effects of crude oil (petroleum) fractions (Meyerhoff, 1975). Therefore this study was undertaken to study the effects of acute toxicity of premium oil (petrol) a fraction of crude oil on periwinkles (*Tympanotonus fuscatus*) an important brackish water specie in the Niger Delta, Nigeria.

MATERIALS AND METHODS

Periwinkles (*Tympanotonus fuscatus*) of size between 4.5 - 5.5cm were handpicked at the Eagle Cement area of the New Calabar River near the Ignatius Ajuru University of Education Rumuolumeni, Port Harcourt. They were transported in plastic buckets to the Chemistry Department Laboratory of the University. Two hundred apparently healthy periwinkles were acclimated to laboratory conditions in plastic tanks of six litre capacity. The tanks were half filled with brackish water and sediments collected from same source. The acclimation was done for seven days. The substrate was prepared by air drying the sediment and then macerated in a mortar and sieved in 2mm mesh.

About 250g of finely prepared sediment were put into each of the plastic tanks to serve as the substrate base. Completely randomized design (CRD) was used for the experiment. The experiment was divided into five treatment levels with three replicates. The test media (petrol) were prepared in the following concentrations: 30.00ml/L, 60.00ml/L, 90.00ml/L, 120.00ml/L, and 150.00ml/L of petrol and a control. Ten of the test animals were introduced into each of the toxicant media. Dead periwinkles were ascertained if the animal has completely retracted into the shell or if it fails to respond to prodding of a glass rod for a period of 15minutes. Mortality assessment was carried out at defined intervals of 24, 48, 72 and 96hours.

The data obtained were subjected to analysis of variance (ANOVA) to determine if significant differences existed between the means in the mortality at different levels of contamination. Where differences existed, Duncan's multiple range test (DMRT) was used to compare the means (Zar, 1984). Toxicological response data involving quantal response (mortality) was analysed using probit analysis (Finney, 1991) to determine the lethal concentrations (LCs) and median lethal times (MLTs).

RESULTS

The total mortality of the periwinkles (*Tympanotonus fuscatus*) in the petrol media indicated a concentration and time dependent death rate. In each of the concentrations, at the different time intervals, the mortality observed were slightly higher than those in the lower concentrations. In the 24^{th} hour, the death rate in 120 and 150 ml/L concentrations were twice as high the value observed in the 30 ml/L concentration. At the 96^{th} hour, the gap in mortality trend tend to close up with deaths recorded in the 30 and 150ml/L which were 22 and 30 respectively (Table 1). The mean mortality of the periwinkle showed slight differences in values between the toxicant concentrations, but significant (P>0.05) variations between the time intervals. However, in the 96^{th} hour, at 150ml/L concentration all the periwinkles died (Table 2).

The lethal effects of the petrol on the periwinkles were expressed as LC_{50} , LC_{90} , LC_{95} and LC_{99} for 48, 72 and 96 hrs. The result showed that there was great variation between the 48th and the 96th hour. The values of the associated lethality decreased progressively as the

exposure duration increased. The 96hr LC_{50} was 34.12ml/L as against the 96hr LC_{99} of 126.64ml/L, while the 48hr LC_{50} was 127.36ml/L and the 48th LC_{99} was 396.82ml/L. The probit equation were Y= -1.10+0.1X, -0.91+0.014X and -0.86+0.25X for 48, 72 and 96 hrs respectively (Table 3).

The MLT₅₀, $_{90, 95}$ and $_{99}$ in the exposure concentrations decreased with time and varied appreciably. The probit equation for the variation were however significant. The MLT₅₀ data obtained with the lower and upper limits were 77.13 (38.50-92.11), 61.64 (30.13-86.41), 68.09 (29.35-75.58), 44.71 (22.98-58.30) and 43.17 (16.95-56.30) hours for 30, 60, 90, 120 and 150ml/L respectively. The MLT₉₉ varied from 164.38 (132.78-260.18) to 101.17 (84.94-122.21) hours between 30.00ml/L to 150.00ml/L of the petrol concentrations (Table 4).

| Time Duration | | | Concentration of kerosene in mg/L | | | |
|---------------|------------------|-----------------|-----------------------------------|-----------------|-----------------|--|
| (Minutes) | 30 | 60 | 90 | 120 | 150 | |
| 24 | 5 ^c | 8^d | 6^d | 10 ^d | 11^d | |
| 48 | 11 ^{bc} | $14^{\rm c}$ | 12° | 15 ^c | $14^{\rm c}$ | |
| 72 | 15 ^b | 19 ^b | 19 ^b | 21 ^b | 24 ^b | |
| 96 | 22 ^a | 25^{a} | 24 ^a | 29 ^a | 30 ^a | |

| Table 1: Total mortality of Tympanotonus fuscatus in different concentrations of petrol after |
|---|
| acute exposure |

Means with the same alphabet in the same column are not significantly different (P>0.05)

 Table 2: Mean mortality of Tympanotonus fuscatus in different concentrations of petrol after acute exposure

| Time Duration Concentration of kerosene in mg/L | | | | | e in mg/L |
|---|----------------------------|--------------------------|----------------------------|-------------------------|--------------------------|
| (Minutes) | 30 | 60 | 90 | 120 | 150 |
| 24 | $1.67\pm0.23^{\rm c}$ | $2.67\pm1.01^{\rm c}$ | $2.00\pm0.00^{\rm c}$ | $3.33 \pm 1.23^{\circ}$ | $3.67 \pm 1.11^{\circ}$ |
| 48 | $3.67\pm0.32^{\text{b}}$ | 4.67 ± 1.74^{bc} | $4.00 \pm 1.25^{\text{b}}$ | $5.00 \ \pm 0.95^{b}$ | $4.67 \pm 1.24^{\rm c}$ |
| 72 | $4.00 \pm 1.10^{\text{b}}$ | $6.33\pm0.55^{\text{b}}$ | 6.33 ± 0.67^{ab} | 7.00 ± 0.00^{ab} | $8.00\pm0.00^{\text{b}}$ |
| 96 | $7.33 \pm 1.32^{\rm a}$ | $8.33 \pm 1.54^{\rm a}$ | $8.00\pm0.00^{\rm a}$ | 9.67 ± 2.01^{a} | 10.00 ± 0.00^{a} |

Means with the same alphabet in the same column are not significantly different (P>0.05)

| able 3: Mean lethal concentration of petro | l to Tympanotonus | <i>fuscatus</i> exposed 96 hours |
|--|-------------------|----------------------------------|
|--|-------------------|----------------------------------|

| Exposure | Lethal conce | Lethal concentrations (ml/L) with associated 95% confidence interval | | | | | |
|-------------------|--------------|--|-----------|-----------|---------------------|-----------|--|
| Duration (hrs) | LC_{50} | LC_{90} | LC_{95} | LC_{99} | Probit equation | Test sig. | |
| 48 | 127.36 | 275.80 | 317.88 | 396.82 | Y= - 1.10+0.1X | *** | |
| 72 | 64.83 | 156.11 | 181.99 | 30.54 | Y= - 0.91+0.014X | *** | |
| 96 | 34.12 | 85.09 | 99.54 | 126.64 | Y= - 0.86+0.25X | *** | |

| Concentration | n Median lethal time (hrs) and associated 95% confidence interval | | | | | | |
|---------------|---|---------------|------------|------------|-------------------|-------------|--|
| of K | МІТ | MIT | MIT | MIT | | π | |
| Kero petrol | MLI_{50} | MLI_{50} | MLI_{50} | MLI_{50} | Probit | Test sig. | |
| (ml/L) | 77.10 | 100.04 | 107.06 | 164.00 | equation | sie sie sie | |
| 30 | //.13 | 122.96 | 137.36 | 164.38 | Y = - | *** | |
| | (38.50- | (101./0- | (113.14- | (132.78- | $1.82 \pm 0.25 X$ | | |
| | 92.11) | 173.90) | 203.27) | 260.18) | | | |
| 60 | 61.64 | 110.82 | 123.89 | 149.68 | Y=- | *** | |
| | (30.13- | (90.09- | | (122.51- | 1.63 + 0.26X | | |
| | 86.41) | 147.50) | (102.41- | 224.50) | | | |
| | | | 173.23) | | | | |
| 90 | 68.09 | 99.48 | 110.07 | 129.96 | Y= - | *** | |
| | (29.35- | (85.56- | (94.65- | (110.30- | 2.13+0.034X | | |
| | 75.58) | 123.73) | 140.81) | 174.29) | | | |
| 120 | 44.71 | 78.02 | 87.46 | 105.17 | Y= - | *** | |
| | (22.98- | (63.85.96.26) | (93.07- | (88.62- | 1.72+0.038X | | |
| | 58.30) | | 109.16) | 135.11) | | | |
| 150 | 43.17 | 75.00 | 84.18 | 101.17 | Y= - | *** | |
| | (16.95- | (60.20- | (69.60- | (84.94- | 1.73+0.04X | | |
| | 56.30) | 93.29) | 105.59) | 122.21) | | | |
| | | | | | | | |
| % | 100 | | | 1 | 1 | | |
| | 90 | | | | | | |
| м | 80 | | | | | | |
| o | 70 | | | | | | |
| r | 60 | | | | Sorios1 | | |
| | 50 | | | | | | |
| a | 40 | | | | Series2 | | |
| | 30 | | | | Series3 | | |
| i | 20 | | | | Series4 | | |
| t | 10 | | | | | | |

Table 4: Median lethal time of petrol with associated 95% confidence interval to Tympanotonus fuscatus after exposure to various concentrations of petrol



90

Concentration of Petrol

120

150

DISCUSSION

y

0

30

60

Crude oil and petroleum fractions cause the blockade of atmospheric oxygen from dissolving in water, thereby limiting the supply of oxygen to aquatic animals, thus resulting in incidences of excretory products in the ambient water environment. The decrease in oxygen content can also explain reasons for mortality of organisms (Ugwu *et al.*, 2011) which in this case is periwinkle. Chemical and physical characteristics of petroleum products dictates the penetrability of the compounds in living organisms and their metabolism and hence the toxic actions they exert on the exposed organism (Chukwu and Odunzeh, 2006). Death of organism can also result from the ability and rate at which the organism excretes transformed products of the chemical which may be more toxic than the parent chemical and or metabolites which may be an all important factor in determining the toxicity of chemicals (Gabriel and Edori, 2010).

Mortality of periwinkles after exposure to crude oil and other petroleum products have been reported (Renner *et al.*, 2008; Ewa-Oboho and Otogo, 2009). The response to death or mortality of the periwinkles was time and concentration dependent similar to that reported in other studies with periwinkles (Chukwu and Odunzeh, 2006; Renner *et al.*, 2008). The 96hr LC_{50} (34.12ml/L) observed indicated that petrol is very toxic to periwinkles. This may have resulted from the volatility of petrol and its penetrating power into the organs of the periwinkle. The interaction of the toxicant (petrol) may have altered the normal biochemistry of the organism to the extent of causing damage. The periwinkle may have adjusted its homeostatic or body defense mechanism to adapt to the new environment up to a point where it got maladjusted thereby leading to death (Gabriel and Edori, 2010).

Death of the organism may have also resulted from respiratory problems, since petrol decreases the oxygen content of aquatic environments and prevents the external organs from interacting with immediate environment by coating the organism. This situation leads to insufficient oxygen supply or utility by organism and therefore suffers from asphyxiation and the organism finally dies.

REFERENCES

- [1]. ATSDR (1999). Toxicological profile for total petroleum hydrocarbons (TPH). Agency for Toxic Substances and Disease Registry, Public Health Service, US Department of Health and Human Services, Atlanta, GA.
- [2]. Carbioch, L., Dauvin, J. C., & Gentil, F. (1977). Preliminary observations on pollution of the seabed and disturbance of sublitoral communities in Northern Brittany by oil from the Amoco Cardiz. *Marine pollution Bulletin*, 9: 303-307.
- [3]. Chukwu, L. O., & Odunzeh, C. C. (2006). Relative toxicity of spent lubricant oil and detergent against benthic macro-invertebrates of a West African estuarine lagoon. *Journal of Environmental Biology*, 479-484.
- [4]. Doertter, J. W. (1992). *Oil spill response in the marine environment*. Pergamon Press, England. 391pp.
- [5]. Ewa-Oboho, I. O., & Otogo, G. A. (2009). Effects of crude oil on the gastropod, *Tympanotonus fuscatus* in the Cross River estuary, South-East Nigeria. *Global Journal of Environmental Sciences*, 8(1): 1-7.
- [6]. Finney, D. J. (1971). *Probit analysis. 3rd edition*. Cambridge University Press, London. pp 5-20
- [7]. Gabriel, U. U., & Edori, O. S. (2010). Quantal responses of hybrid catfish (*Heterobranchus bidorsalis* x *Clarias gariepinus*) to agrolyser. *Journal of League of Researchers in Nigeria*, 11(1): 67-72.
- [8]. Howard, I. C., Gabriel, U. U., & Horsfall, M. (2009). Evaluation of total hydrocarbon levels in some aquatic media in an oil polluted mangrove wetland in the Niger Delta. *Applied Ecology and Environmental Research*, 7(2): 111-120.
- [9]. Jee, J. H., & Kang, J. C. (2005). Biochemical changes of enzymatic defense system after phenanthrene exposure in olive flounder, *Paralichthys olivaceus*. *Physiological Research*, 585-591.

- [10]. Law, R. J., & Biscaya, J. L. (1994). Polyaromatic hydrocarbons (PAH) Problems and progress in sampling, analysis and interpretation. *Marine Pollution Bulletin*, 29: 235-241.
- [11]. Lopes, P. A., Pinihiro, M. C., Santtos, M. L., Mathias, M. J. Collares, P., & Vegas-Crepo, A. M. (2001). Responses of antioxidant enzyme in fresh water fish pollutions (*Leucisus alburnoides* complex) to inorganic pollutants exposure. *Scientific Total Environment*, 280: 153-163.
- [12]. Medeiros, P. M., Bicego, M. C., Castelao, R. M., Rosso, C. D., Fillmamm, G., & Zamboni, A. J. (2005). Natural and anthropogenic hydrocarbon inputs to sediments of Patos Lagoon Estuary. *Brazil Environment International*, 31: 77-87.
- [13]. Meyerhoff, R. D. (1975). Acute toxicity of benzene, a component of crude oil to juvenile striped bass (*Morone saxatillis*). Journal of Fisheries Research Board Canada, 32:1864-1866.
- [14]. Oluah, N. S. (2001). The effect of sublethal cadmium on the haematology of the freshwater fish, *Clarias gariepinus* (Pisces: Clariidae). *Journal of Science of Agriculture, Food Technology and Environment, 1*:15-18.
- [15]. Patin, S. (1999). Environmental impact of the offshore oil and gas industry-Ecomonitor East Northport New York, 425pp.
- [16]. Renner, K. O., Don-Pedro, K. N., & Nubi, O. A. (2008). Oil spillage and its impact on the edible mangrove periwinkle, *Tympanotonus fuscatus* Var *Radula* (L). *Science World Journal*, 3(3): 13-16.
- [17]. Ugwu, L. L. C. Ude, E. F., Nwamba, H. O. and Chima, I. N. (2011). Effect of crude oil and some petroleum products on *Clarias gariepinus* fingerlings (catfish Clariidae). Continental Journal of Fisheries and Aquatic Science, 5(1): 24-30.
- [18]. Widdows, J., Bakke, T., Bayne, B. L., Donkin, D. R., & Lowe, D. M. (1982). Response of *Mytilus edulis* on exposure to the water-accommodated fraction of North Sea oil. *Marine Biology*, 67: 15-31.
- [19]. Zar, H. K. (1984). *Statistical tools for scientific analysis*. Oxford Publishers, London, 319pp.